

CALCULATION OF THE SHEDDING RATE OF CRYPTOSPORIDIUM OOCYSTS FROM THE NATURAL INFECTED SHEEP

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ABSTRACT

The objective of this study to calculate means of oocysts in feces natural infected sheep. A total of 150 sheep fecal samples collected from different regions of Baghdad province (AL-Sholla, AL-Horia, AL-Taji, and Abo-Grab)-Iraq, during the period from the beginning of January to the end of May 2016. Three laboratory methods (modified acid fast stain, sheather's sugar solution and calculation of oocysts in feces of infected animals by haemocytometer) were used for diagnosis of Cryptosporidial oocysts and study the shedding rate of oocysts from infected male and female (pregnant and non pregnant) animals. The total infection rate was 44.66% (67/150), and the highest infection rate was detected in March, while the lowest infection rate was recorded in January 63% (19/30), 20% (6/30) respectively. The results were revealed significance difference between male and female infection rate, 31.74% (20/63), 54.02% (47/87) respectively. The average number of shed oocysts per gram of feces from infected non pregnant ewes was 1440 oocysts per gram while in infected pregnant ewes was 2082 oocysts per gram. The shedding rate of oocysts in pregnant ewes show the highest rate in the end period of pregnancy, while the lowest rate of shedding in the beginning period of pregnancy.

Keywords: *Cryptosporidium*, oocysts, shedding, pregnant, sheep

كوان

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حساب معدل طرح اكياس بيض البوغ الخبيء من الاغنام المصابة طبيعيا

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المستخلص

كان الهدف من الدراسة حساب معدل طرح اكياس بيض البوغ الخبيء من الاغنام المصابة طبيعيا حيث تم جمع 150 عينة براز اغنام تضمنت مناطق مختلفة من محافظة بغداد | العراق (الشعلة، الحرية، التاجي، وابو غريب) اعتبارا من شهر كانون الاول الى شهر مايس 2016. استعملت ثلاث طرق مختبرية (صبغة زيل نلسون المحورة) محلول شيندر السكري و حساب اعداد اكياس البيض في البراز بواسطة الهيموسايتوميتر (لتشخيص اكياس بيض الطفيلي ودراسة معدل طرح اكياس البيض من الذكور والاناث (الحوامل وغير الحوامل) المصابة. اعلى نسبة للإصابة الكلية بلغت (67/150) 44.66%، وسجلت اعلى نسبة للإصابة في شهر آذار واوطأها في كانون الثاني وبلغت (30/19) 63% و (30/6) 20% على التوالي، سجلت الدراسة فرقا احصائيا في نسب الإصابة بين الذكور والاناث، (63/20) 31.74% و (87/47) 54.02 % على التوالي كان معدل طرح اكياس البيض لكل غرام براز من النعاج غير الحوامل المصابة 1440 كيس بيضة لكل غرام. في حين بلغ 2082 كيس بيضة لكل غرام في النعاج الحوامل . سجل اعلى معدل طرح اكياس البيض للنعاج الحوامل في الفترة الاخيرة من الحمل، في حين سجل اقل معدل للطرح في بداية فترة الحمل.

الكلمات المفتاحية: البوغ الخبيء، اكياس البيض، طرح، حمل، الاغنام

INTRODUCTION

Cryptosporidium is an intestinal protozoan parasite that allegedly infects alimentary canal of human and animals, it considered as one of important zoonotic parasite (21). *Cryptosporidium* was first described impacting in rats by Tyzzer in 1907 (40). Cryptosporidiosis was first described in lamb with diarrhea in Australia in 1974 by Barker and Carbonell and has consequently been revealed in 12 other nations (21). Younger animals (calves, lambs, goat kids) appear to be more delicate to sickness, in neonatal ruminants, cryptosporidiosis considered a main cause of diarrhea and death, with important economic loss (2, 14, 19, 23, 28, 38, 40). The parasite has a monoxenous life cycle which means asexual and sexual stages happen within one host, within 1-3 days, the prepatent period varies from 1-3 weeks, whereas the patent period which means duration of oocysts shedding, can differ from several days to month, indicating the possibility of this infection continue to persist. (31). Cryptosporidiosis transmitted via the fecal-oral transmission from infected animal to the another or from infected animals to human through contaminated food and water with mature oocysts (21, 32). The prevalence of *Cryptosporidium* infection in different reports reach to 85% in lambs, some of these reports, in Ethiopia, 2.6%, in Australia 3.7%-47%, in Brazil 13.6%-46.5%, in Turkey 25.7% in Mexico, 29%, and 42.1% in Serbia (32, 37). In Iraq the parasite recorded by Al-Zubaidi, (8) and Al-Azzui, (4) in cattle, while in sheep and lambs the parasite recorded by Abd-Alwahab, (3), Kadhim, (24), Al-Seady, (7) and Al-Zubaidi, (10). *Cryptosporidium* considered as the most important parasite causing diarrhea in suckling lambs, and adult ewes act one of the main sources of infection of these lambs. This study aimed to investigate the shedding rate of

Cryptosporidium oocysts from natural infected sheep.

MATERIALS AND METHODS

A total of 150 sheep fecal samples were collected from both sexes (87 females and 63 males) from different regions of Baghdad province (AL-Sholla, AL-Horia, AL-Taji, and Abo-Grab) -Iraq, during the period from January to May 2016. Fecal samples were collected directly from the rectum of animals, in a clean plastic container and given sequential numbers, and all information, age, sex and date of sampling were recorded on it. The samples were transported in a cooling box to the department of parasitology in the College of Veterinary Medicine-University of Baghdad for laboratory diagnosis.

Examination of samples: Three laboratory methods were used to diagnose *Cryptosporidium* oocysts, and study the shedding rate of oocysts from infected male and female (pregnant and non pregnant), Sheather's sugar solution (Fig: 1) Modified Zeihl-Neelsen Stain (MZN) (Fig: 2) and calculation of *Cryptosporidium* oocysts in feces of infected sheep by using haemocytometer. (1, 10, 12, 13, 16, 41).

Isolation and Calculation of the *Cryptosporidium* oocysts

After isolating and purifying the parasite oocysts which found in the feces of infected sheep by using flotation with Sheather's sugar solution (4, 9, and 16). The number of oocysts calculated in 1 ml of suspended oocysts solution by using haemocytometer slide which used for white blood cells calculation in the eight squares of the two chamber of this slide, then the total number of oocysts per 1ml calculate according to the following equation: Al-Attar, (1) (Fig: 1)

Number of oocysts in 1 ml = (1000 x calculated oocysts number) / 8

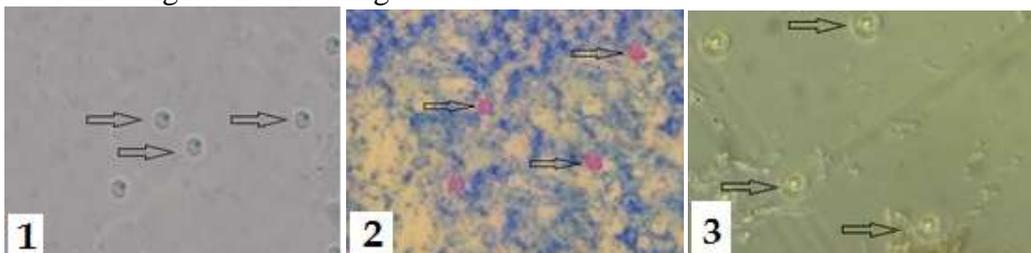


Fig 1. *Cryptosporidium* oocysts in Flotation by sheather's sugar solution x10

Fig 2. *Cryptosporidium* oocysts by modified acid fast stain x100

Fig 3. Calculation of *Cryptosporidium* oocysts by haemocytometer slide 40x15

Statistical analysis: The Chi-square test was used for the comparison between the results. One way ANOVA was also performed and comparisons were done using LSD. Differences were considered statistically significant at $P < 0.05$ (38).

RESULTS AND DISCUSSION

The results of this study recorded 44.66 % (67/150) of sheep infected with Cryptosporidiosis in different areas of Baghdad city, the highest infection rate in AL-Taji 60 % (21/39), while the lowest rate of infection in Abo-Grab 32.5% (13/36) (Table:1). This results agreed with Khalil, (26) in Mosul and Al-Gilani, (5) in Baghdad who recorded infection rate in sheep 36.43%, 48.8% respectively, also agreed with Al-Kaabi, (6) in Diwaniyah province who found 27.5% of sheep shed *Cryptosporidium* oocysts, also the result agreed with Faleket *et al*,

(18) in Nigeria who recorded 22.7 of sheep infect with Cryptosporidiosis. while the result disagreed with Abd-Al-Wahab, (3) who found higher infection rate 74.2% and Khadim, (24) who show lower infection rate 15.8 in Baghdad city, also disagree with Nouri and Karami, (30) and Harandi and Ardakani, (23) in Iran who recorded 17.2%, 13.3% of sheep respectively infected with cryptosporidiosis, and also disagreed with Rayan *et al*, (34) in Australia and Tembue *et al*, (39) in Brazil who found 2.6% , 3.7% of sheep respectively infected with the parasite. The differences in infection rates in different regions in the world and in Iraq may be due to differences in the number of animals checked, sample size, climatic conditions, and as well as different diagnostic methods used in laboratory. (21)

Table 1: Prevalence of *Cryptosporidium* spp according to areas

| Areas | No. of Examined fecal Samples | No. of positive fecal samples | percentage % | Chi square value | P |
|-----------|-------------------------------|-------------------------------|--------------|------------------|------|
| AL-Sholla | 35 | 17 | 43.58 | 2.96 | 0.39 |
| AL-Horia | 40 | 16 | 44.44 | | |
| AL-Taji | 39 | 21 | 60 | | |
| Abo-Grab | 36 | 13 | 32.5 | | |
| Total | 150 | 67 | 44.66 | | |

The result of study indicated that the animals age effect on infection rates with the highest rate of infection showed in age group 6-12

months 71.42% (21/35) while the lowest infection rate recorded in age group < 36 months 30% (12/40), with significant differences at $p < 0.05$ (Table: 2).

Table 2. Prevalence of *Cryptosporidium* spp according to the age groups

| Age (months) | No. of Examined fecal Samples | No. of positive fecal samples | percentage % | Chi square value | P |
|--------------|-------------------------------|-------------------------------|--------------|------------------|------|
| 6-12 | 35 | 21 | 71.42 | 7.22 | 0.06 |
| 13-24 | 36 | 15 | 41.66 | | |
| 25-36 | 39 | 15 | 38.46 | | |
| 36< | 40 | 12 | 30 | | |
| Total | 150 | 67 | 44.66 | | |

This results agreed with Abd-Alwahab, (3), Kadhim, (24) and Al-Zubaidi, (10) in Baghdad who found the highest infection rate of cryptosporidiosis in age group less than six month which reach 81.46%, 34.95% and 70% respectively. Also the result agreed with Xiao *et al*, (42), Causpeet *et al*, (15) who recorded high infection rate in neonatal lambs 78.3% and 66.4% respectively. This finding agreed

with EI-Wahed (17) in Egypt, Sari *et al*. (35) in Turkey who reported high prevalence rate of parasite in small lambs. The highest infection rate in small animal may be due to high shedding rate of *Cryptosporidium* oocysts from dam which contaminate the food and water in farm and increase the chance to infect lambs (21). The result of this study showed significance difference ($p < 0.05$) in infection rate according to the months of study, the

highest rate of infection recorded in March 63.33% (19/30), while the lowest rate of infection recorded in January 20% (6/30) (Table: 3). This result agrees with Abd Al-Wahab (3) and Kadhim (24) and Al-Zubaidi, (10) who recorded high infection rate of cryptosporidiosis among lambs in March and April. This re-

sult may be due to good environmental condition (temperature and humidity) for the parasite and large number of *Cryptosporidium* oocysts, that shed from pregnant and lactating ewes in the farm which considered as a source of infection to the lambs (21,22,27,35)

Table 3. Prevalence of *Cryptosporidium* spp according to the Months

| Examination methods | No. of Examined fecal Samples | No. of positive fecal samples | percentage % | Chi square value | P |
|---------------------|-------------------------------|-------------------------------|--------------|------------------|-------|
| January | 30 | 6 | 20 | 13.64 | 0.008 |
| February | 30 | 11 | 36.66 | | |
| March | 30 | 19 | 63.33 | | |
| April | 30 | 15 | 50 | | |
| May | 30 | 16 | 53.33 | | |
| Total | 150 | 67 | 44.66 | | |

The study shows significance difference between male and female infection rates, 31.74% (20/63) and 50.02% (47/87), respectively (Table 4). This result agreed with Akinkuotu and Fagbemi, (2014) in Nigeriawho found the high infection rate in females than males, 48.4%, 30.4% respectively, while this result disagreed with Abd Al-Wahab (3), Kadhim (24) in small lambs in Baghdad city and Rasheed (33) in goat kids in Iraq, who found no significance differences in the infection rate between male

and female due to equal possibility of exposure to the contaminated environment (20). Also Table (4) shows the highest infection rate recorded in pregnant ewes while the lowest rate in non pregnant ewes, 72.5% (29/40), 38.29% (18/47) respectively, with significant differences ($p < 0.05$) this result may occur due to the hormonal changes in pregnant animals and its effects on the immune status of the animal body (21,28,43).

Table 4. Prevalence of *Cryptosporidium* spp according to the sex and status

| Sex | No. of Examined fecal Samples | No. of positive fecal samples | percentage % | Chi square value | P |
|--------------|-------------------------------|-------------------------------|--------------|------------------|-------|
| Male | 63 | 20 | 31.74 | 7.33 | 0.006 |
| Female | 87 | 47 | 54.02 | | |
| Pregnant | 40 | 29 | 72.5 | 10.17 | 0.001 |
| Non pregnant | 47 | 18 | 38.29 | | |

The study Calculate the shedding rate of *Cryptosporidium* oocysts from natural infected sheep, male and females (pregnant and non pregnant) (Table: 5). The increase and decrease of oocysts shedding in feces of infected animals occurring due to the hormonal changes and its effects on the immune status of the animals (11,21,28). The results of study recorded the highest mean numbers of oocysts shedding per gram of feces seen in pregnant ewes than non pregnant, 2082 oocysts per/gm 1440 oocysts per/gm respectively **with** significant differences, and also the highest mean numbers of oocysts shedding per gram was shown in

the end period of pregnancy while the lowest number of oocysts shedding per/gm recorded in infected males, 2460 oocysts per/gm, 1020 oocysts per/gm respectively with significant differences. This results agreed with Kehrli *et al.*, (25) and Nckerson *et al.*, (29) who recorded a decreasing in efficiency of neutrophil cells during pregnancy, perparturition, parturition and post parturition periods, also Yang *et al.*, (44) who recorded a decreasing in B and T lymphocyte cells 60% and 40% respectively in same period, which leads to reduced the immunoglobulin's especially the major types, IgG, IgM and IgA, synchronized with the pe-

riod of parturition and lactation, that lead to increase of oocysts shedding per/gm from infected animals.

Table 5. Total No and mean of oocysts shedding from infected animals

| Sex of animals | Infected animals | Total No of shed oocysts per/gm feces | Mean No of shed oocysts per/gm feces |
|---------------------|------------------|---------------------------------------|--------------------------------------|
| Male | 20 | 28560 | 1020±88.67 ^c |
| Non pregnant female | 18 | 20160 | 1440±98.65 ^d |
| Pregnant Female | 29 | 52050 | 2082±211.43 ^b |
| First period pre. | 15 | 27450 | 1830±187.66 ^c |
| End period pre. | 10 | 24600 | 2460±223.21 ^a |
| Total | 67 | 100770 | 1504±114.21 |

Different superscript refers to significant differences at $p < 0.05$

In conclusion, the increase of mean number of oocysts shedding in feces of infected pregnant ewes than males which occurring due to the hormonal changes and its effects on the immune status of the animals.

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REFERENCES

- Al-Attar, M. A. 1981. Factors Affecting the Pathogenesis of *Eimerianicatrix* Infections in Ckicken. Ph.D. Dissertation.University of Guelph, Canada.pp:21-23
- Akinkuotu, O.A.,and B.O.Fagbemi.2014. Occurrence of *Cryptosporidium* species coproantigens on a University Teaching farm in Nigeria.Sokoto Journal of Veterinary Sciences, 12 (2):41-46
- Abd Al-Wahab, I.H. 2003. Study in the epidemiology of the Intestinal Protozoa (*Eimeriaspp.*, *Cryptosporidium spp.*, *Giardia spp.*) in the Sheep in Baghdad Province. M.Sc. Thesis, College of Veterinary Medicine, University of Baghdad.pp:123.
- Al-Azzawi, M.H.K. 2003. Epidemiological Study of Cryptosporidiosis and the Isolation of the Parasite Antigens, Diagnosis and the use of some Medicinal Plant Extracts Work Therapy. Ph.D. Dissertation.. College of Vet. Med. University of Baghdad.Iraq. pp: 157
- Al-Gilani, Abdul Wahab 2003.Epidemiological and Diagnostic Study of *Cryptosporidium* in Humans and Animals in the Village of White Gold. Ph.D.Dissertation, College of Veterinary Medicine, University of Baghdad.pp:107
- Al-Kaabi, P. R. 2005. Epidemiological study of *Cryptosporidium* oocysts as causes of Diarrhea in Diwanayah Province. M.ScThesis. Faculty of veterinary medicine, University of Qadisiya.pp:109
- Al-Seady, H.H.O. 2014. Diagnostic and Epidemiological Study of Intestinal Protozoa Causes of Diarrhea in Lambs in Baghdad Province. M.ScThesis.Collegeof.Vet.Med.Baghdad University, Iraq.pp:122
- AL-Zubaidi, M.Th.S. 1994. Cryptosporidiosis in Calves. M.Sc. Thesis. Faculty of Veterinary Medicine, University of Baghdad.pp:102
- AL-Zubaidi, M.Th.S. 2009.Some Epidemiological Aspects of Cryptosporidiosis in Goats and UltrastructuralStudy Ph.D. Dissertation., University of Baghdad. pp:133
- Al-Zubaidi, M.Th. S. 2017.Molecular and microscopic Detection of *Cryptosporidium spp* in Sheep in AL-Taji area-Baghdad/Iraq. .International Journal of Science and Nature. 8 (2) :372-376
- Atwill, E. R., B. Hoar, M. das Gracias Cabral Pereira, K. W. Tate, F. Rulofson, and G. Nader. 2003. Improved Quantitative Estimates of Low Environmental loading and Sporadic Periparturient Shedding of *Cryptosporidium parvum* in Adult Beef Cattle.Appl Environ Microbiol., 69:(8)4604–4610
- Baxy,D.;Blundell,N.and C.A Harf.,(1984) The development and performance of a simple sensitive methods for the detection of *Cryptosporidium* oocysts in feaces. J. Hyg. 23:317-323
- Beaver, P.C. and R.C Jung.1985. Animal Agents and Vectors of Human Diseases .(5th ed.) Lea and Febiger .pp: 249

14. Bhat S.A., P.D.Juyal, N.K.Singh, L.D.Singla 2013. Coprological investigation on neonatal bovine cryptosporidiosis in Ludhiana, Punjab. *Journal of Parasitic Diseases*, 37(1): 114-117
- 15-Causape,A.C.,J.Quilez,C.Sanchez-Acedo, E. Delcachoand L. F.Bernad.2002. Prevalence and analysis of potential risk factors for *Cryptosporidium parvum* infection in lambs in Zaragoza (northeastern Spain). *Vet. Parasitol.*104(4):287-298
16. Chermette, R., and Q. S. Boufassa,. 1988. Cryptosporidiosis a Cosmopolitan Disease in Animals and Man , 2nd ed . Office International Epizooties. France.pp: 122
17. El.Wahed,M.A.1999.*Cryptosporidium* infection among sheep in Qalubia governorate.Egypt.J.Egy.Soc.Parasitol.29:113-118.
18. Falek, O.O.,K.Sahabi,,and A.B.Aliyu.2006. Prevalence of *Cryptosporidium* in slaughter sheep and goats at Sokoto,Nigeria .*Anim. Produ. Res.Adv .* 2(3): 179-182
19. Fayer, R. 2010. Taxonomy and Species delimitation in *Cryptosporidium*.*Experimental Parasitology.* 124 (1): 90-97
20. Fayer,R.,M.Santin,,and J.M. Trout. 2008. *Cryptosporidium ryanaen.sp .*(Apicomplexa: cryptosporidii) in cattle(*BosTaurus*). *Vet.Parasitol.*156 (3-4):191-198 .
- 21-Fayer, R. and L. Xiao, 2008. *Cryptosporidium* and Cryptosporidiosis. 2nd ed. CRC.Press. pp: 560
22. Giadinis N.D, S Symeoudakis, E, S.Q.Lafi, and H.Karatzias. 2012. Comparison of two techniques for diagnosis of cryptosporidiosis in diarrhoeic goat kids and lambs in Cyprus. *Tropical Animal Health and Production*, 44(7): 1561-1565
23. Harandi, F.M.and F.R. Ardakani. 2008. *Cryptosporidium* infection of sheep and goats in Kerman:Epidemiology and risk factor analysis.*J.Vet .Res.* 63 (1):47-51
24. Kadhim, Th.A. 2009. Epidemiological and Histopathological Study of Cryptosporidiosis in Sheep in Baghdad Province M.Sc. Thesis. Faculty of Veterinary Medicine, University of Baghdad.pp:89
25. Kehrli, M. E.,B.J.Nnnecke, and J.A.Roth,. 1989. Alteration in bovine peripheral blood neutrophil function during the peripartum period.*Am.J.Vet.Res.*50:207-214
26. Khalil, L.Y. 2000.Compared the Efficiency of Some Diagnostic Tests for Cryptosporidiosis in Lambs and Kids in Nineveh. M.Sc. Thesis – College of Veterinary Medicine, University of Mosul.pp:78
27. Maurya, P.S., R.L.Rakesh, B.Pradeep,S.Kumar, K.Kundu, R.Garg, H.Ram, A.Kumar, P., S.Banerjee. 2013. Prevalence and risk factors associated with *Cryptosporidium* spp. infection in young domestic livestock in India. *Tropical Animal Health and Production*, 45(4): 941-946
28. Mallard, B.A., Dekkers, J.C., M. J.Ireland,K.E.Leslie,S.Sharif, Lacey,C Wagher,L. and C.Wilkie. 1997. Alteration in Immune Responsiveness During the peripartum period and its ramification on dairy Cow and Calf Health.*J.Dairy.Sci.*81:585-595
29. Nckerson,S.C., W.Owens,J.J.Rejman, and S.P.Oliver. 1993. Effect of interleukin-1 and interleukin-2 on mammary gland leukocyte population and histology during the early non lactating period.*Zntralbl. Veterinaermed. ReiheB.*40 :621-633
30. Nouri,M.and M. Karami,.1991. Asymptomatic Cryptosporidiosis in nomadic shepherds and their sheep.*J.Infec.*23(3):331-333
31. O'Donoghue, P. N. J.1995. *Cryptosporidium*andcryptosporidiosis in man and animals.*Int.J.Parasitol.*25:139-195.
- 32-Ozdal, N., P.Tanritanir,Y.Goz,S.Deger, and S.Kozat.2009. Parasitic protozoans (*Eimeria, Giardia, and Cryptosporidium*) in lambs with diarrhoea in the Van province, Turkey. *Bull. Vet. Inst. Pulawy*, 53: 47-51
- 33-Rasheed, R.N.1997. Cryptosporidiosis in Iraqi goat kids. *The Veterinarian.* 1(6):1-5
- 34.Ryan,U.M.,C.Bath,I.Robertson,C.Read,A.E lliot,L.Mcinnis,R.Trub,,andB.Besier.2005. Sheep may not be an important zoonotic reservoir for *Cryptosporidium* and *Giardia* parasites.*Appl.Enviro.Microbiol.*71(9):4992-4997
- 35-Sari, B.,M.O.Arslan,Y.Gicik, M. Kara, and G.T.Tasci,. 2008. The prevelance of *Cryptosporidium* species in diarrhoeic lamb in Kars province and potential risk factor. *Trop. Anim.Health Prod.*, 41(5):819-26
- 36-Santín, M. 2013.Clinical and subclinical infections with *Cryptosporidium* in animals. *N. Z. Vet. J.*, 61: 1-10
37. Silva- Fiuza, V. R., R.I.Juliboni-Cosendey, E. Frazao-Teixeira, M. Santin, S. Pedraza- Di-

- az, C. Amar, G.L. Nichols, and J. McLauchlin. 2001. Nested polymerase chain reaction for amplification of the *Cryptosporidium* oocyst wall protein gene. *Emerging Infectious Diseases*. 7:49–56
38. Snedecor, G.W., and W.G. Cochran, 1989. *Statistical Methods*, Eighth Edition, Iowa State University Press
39. Tembue, A. A., M. Alves, J. Borges, C. G. Faustino, M. A. DaGloria. And E.L.C. Machado. 2006. *Cryptosporidium* spp. in sheep in Ibimirim district, Pernambuco state, Brazil. *Cienc. Vet. Trop. Recife-PE*. 9(10):41-43
40. Xiao, L. 2010. Molecular epidemiology of cryptosporidiosis: an update. *Experimental Parasitology*, 124 (1): 80–89
41. Xiao, L., R.U. Fayer, D. M. Ryan, and S.J. Upton, 2004. *Cryptosporidium* taxonomy: recent advances and implications for public health. *Clin Microbiol. Rev.*, 17(1):72–97.
42. Xiao, L., R.P. Herd, and D.M. Rings. 1993. Diagnosis of *Cryptosporidium* on a sheep farm with neonatal diarrhea by immunofluorescence assay. *Vet. Parasitol.*; 47 (1-2) : 17-23
43. Yui, T., T. Nakajima, N. Yamamoto, M. Kon, N. Abe, and M. Matsubayashi. 2014. Age-related detection and molecular characterization of *Cryptosporidium suis* and *Cryptosporidium scrofarum* in pre- and post-weaned piglets and adult pigs in Japan. *Parasitology Research*, 113(1):359-365
44. Yang, T.J., J.F. Mather, and E. D. Rbinovsky. 1988. Changes in subpopulation of lymphocytes in peripheral blood and supra mammary and pre scapular lymph nodes in cows with mastitis and normal cows. *Vet. Immuno. Immunopathol.* 18:279-285.